

Analysis To Support Allergen Risk Management: Which Way To Go?

Tatiana Cucu, Liesbeth Jacxsens, and Bruno De Meulenaer*

NutriFOODchem Unit (member of Food2Know), Department of Food Safety and Food Quality, Ghent University, Coupure Links 653, B-9000 Gent, Belgium

ABSTRACT: Food allergy represents an important food safety issue because of the potential lethal effects; the only effective treatment is the complete removal of the allergen involved from the diet. However, due to the growing complexity of food formulations and food processing, foods may be unintentionally contaminated via allergen-containing ingredients or cross-contamination. This affects not only consumers' well-being but also food producers and competent authorities involved in inspecting and auditing food companies. To address these issues, the food industry and control agencies rely on available analytical methods to quantify the amount of a particular allergic commodity in a food and thus to decide upon its safety. However, no "gold standard methods" exist for the quantitative detection of food allergens. Nowadays mostly receptor-based methods and in particular commercial kits are used in routine analysis. However, upon evaluation of their performances, commercial assays proved often to be unreliable in processed foods, attributed to the chemical changes in proteins that affect the molecular recognition with the receptor used. Unfortunately, the analytical outcome of other methods, among which are chromatographic combined with mass spectrometric techniques as well as DNA-based methods, seem to be affected in a comparable way by food processing. Several strategies can be employed to improve the quantitative analysis of allergens in foods. Nevertheless, issues related to extractability and matrix effects remain a permanent challenge. In view of the presented results, it is clear that the food industry needs to continue to make extra efforts to provide accurate labeling and to reduce the contamination with allergens to an acceptable level through the use of allergen risk management on a company level, which needs to be supported inevitably by a tailor-validated extraction and detection method.

KEYWORDS: food allergens, risk management, detection, challenges, food processing, extraction, competent authority, food company

■ INTRODUCTION

Food allergies involve abnormal responses to specific foods (mostly proteins) that are normally harmless and are mediated by the immune system.¹ Food allergens pose a risk only to a limited number of consumers while being harmless to most other consumers regardless of the amount ingested. When ingested by allergic consumers, the symptoms can range from mild to severe and life-threatening.² Food allergies are estimated to affect about 2% of the adult population in industrialized countries, and their prevalence is reported to be higher in infants and children (6–8%).^{3,4} Over 180 allergenic food proteins have been identified to date, with a few major allergens occurring in common foods (e.g., egg, milk, fish, crustaceans, peanut, soybean, wheat, and tree nuts).⁵ Food allergens are almost always proteins or glycoproteins with molecular weights of 5–70 kDa.³ They mostly represent the major protein fraction of a particular allergenic food commodity and are reported to be typically resistant to proteolysis and stable during food processing.

It is well reported that for very sensitive patients trace amounts of allergens can induce severe and even fatal reactions. For example, as little as 30 μg of hazelnut is able to elicit an allergic reaction,⁶ whereas the predicted threshold value giving a one-in-a-million response rate was reported to be 0.07 μg of milk, 0.003 μg of egg, 0.5 μg of peanut, and 0.3 mg of soybean.⁷ Therefore, the only effective treatment for food allergies is their complete avoidance from the diet.⁸ Food allergens represent a serious safety issue because many of the allergic food commodities are important nutrient sources (milk, eggs, wheat-based products, etc.), and thus their complete exclusion

from the diet is not possible or desirable. Moreover, because of their functionality, several of these products or products thereof are frequently used as ingredients in various composite foods. For individuals affected by severe, life-threatening food allergies, this is a significant food safety issue and thus the protection of such allergic consumers is of concern for the food industry and public health authorities. Therefore, the food industry is obliged to provide accurate labeling by clearly indicating the composition of each food product and to bring safe products on the market. The European Commission (Directive 2007/68/EC) set up a list of allergens that have to be labeled on foods regardless of the amount deliberately added as ingredient⁹ (Table 1). In various countries, similar lists and labeling obligations are applicable.^{10–12} Unfortunately, despite these legislative frameworks, food allergens can still inadvertently be present in a product mainly due to the fact that several different food products are made within the same plant, which can lead to an in-process and postprocess cross-contamination. Cross-contamination might be caused by improper equipment cleaning/sanitation procedures, in the case of a change from one product to the next, but also due to rework.¹³ This leads to the presence of the so-called "hidden allergens". Over 600 alerts due to the presence of undeclared allergens in foods have been

Special Issue: Advances in Food Allergen Detection

Received: July 30, 2012

Revised: January 15, 2013

Accepted: January 16, 2013

Published: January 16, 2013

Table 1. Annex III a of Directive 2007/68/EC

cereals containing gluten and products thereof
crustaceans and products thereof
eggs and products thereof
fish and products thereof
peanuts and products thereof
soybeans and products thereof
milk and products thereof
nuts and products thereof
celery and products thereof
mustard and products thereof
sesame seeds and products thereof
lupin and products thereof
molluscs and products thereof
sulfur dioxide and sulfites (concentrations of >10 mg/kg or 10 mg/L)

reported by the Rapid Alert System for Food and Feed in the European Union (EU) alone.¹⁴ The presence of such hidden allergens poses a serious threat to the allergic consumer and might lead to food recalls, which are expensive for the industry.¹⁵ Moreover, in the European “General Food Law” it is stated that food manufacturers are responsible for the safety of food products brought onto the market.¹⁶ This means that food manufacturers need to take extra measures to prevent or control cross-contamination to protect the allergic consumers and their own reputations. Because no specific legislative framework is available for the labeling of food products possibly contaminated with allergens, it is not always clear for food producers how to manage this issue. Therefore, warning labeling messages such as “May contain ...” or “Present in the processing environment” or “This product is made on a line/in a factory that also handles...” are often used. However, the extensive use of such warning messages can be confusing for the allergic consumers and in parallel leads to a narrowing of the food products available for them. Eating outdoors is another issue for the allergic consumers, indicating that they are forced to rely mainly on the food prepared at home. To offer some comfort to allergic consumers, warning messages should be accurately applied. This implies that they are supported by a proper risk management on company level. Pele et al.¹⁷ showed that food products free of warning messages are often contaminated with food allergens, whereas some of the labeled foods were reported to be allergen free. This indicates that such extensive labeling practices run the risk of undervaluing the labels, and as a further consequence consumers lose their trust in them and in food producers, applying to all of them a negative perception.

To provide accurate information for allergic consumers, the food industry must have access to reliable extraction and detection methods, meaning tailored and validated to the production and product characteristics, and insight into possible contamination routes during processing and manufacturing. Such extraction and detection methods are needed to screen the incoming materials for the absence of undeclared residues of allergens, to evaluate the efficiency of the implemented preventives measures such as sanitation programs applied to remove residues of allergenic foods from shared equipment in the framework of risk management at the company level, and to control the end products. Moreover, the availability of robust analytical methods would allow officials to decide whether a product should be recalled or not in the case of calamities. An allergen risk management program at the

company level to control the presence of hidden allergens, backed up by reliable extraction and detection methods, validated for the company-specific production processes and product matrix, should be part of the food safety management system in a food company.

■ QUANTIFICATION OF FOOD ALLERGENS

Currently there are several analytical approaches applied for the detection and quantification of allergen traces in food products.^{3,18–20} These can either target the allergen itself (one or several proteins) or a marker that indicates the presence of the allergenic commodity. Among the methods targeting the allergen (protein-based methods), the most used are the receptor-based methods (e.g., antibody based enzyme-linked immunosorbent assays (ELISA) and biosensors^{21–27} and non-antibody based such as DNA aptamers^{28,29}) and chromatographic and mass spectrometric methods. DNA-based methods such as polymerase chain reaction (PCR) with real-time PCR providing quantitative results are also available for food allergen detection but are an indirect indicator of the potential presence of the targeted allergen.

There are a number of requirements for the methods used for allergen determination in food. The used analytical methods need to be specific for the targeting compound, highly sensitive so that preferably the lowest amount able to trigger an allergic reaction can be detected, must be specific, and should not be influenced by the presence of matrix components so that false-positive and false-negative results are avoided. Only recently did protocols on the validation of commercially available allergen detection systems become available by the European Committee for Standardization (CEN).^{30–32}

Protein-Based Methods. Receptor-Based Methods. Receptor-based methods are widely used for allergen detection and are based on the interaction between specific antibodies and an antigen (food allergen). They can be developed to detect one or several proteins (allergenic or not) from the allergenic food commodity. Because foods contain several proteins, the choice of the appropriate analyte is crucial for the development of reliable methods. Another important aspect for the receptor-based methods is the choice of the primary antibodies (monoclonal or polyclonal). Nowadays, ELISAs are the method of choice by the food producers and control agencies for routine analysis of food allergen contaminations, and commercially available kits are mostly used. ELISA is used because it is relatively cheap and easy to perform and has a high sensitivity. Generally, two formats of ELISA are available: sandwich and competitive. In the case of the sandwich ELISA the primary antibody immobilized on the plate captures the food allergens, which are further detected by a secondary allergen-specific antibody labeled with enzyme. In the case of competitive ELISA, food allergens immobilized on the plate are competing with the allergens from the sample to bind with the primary antibody labeled with enzyme. Unfortunately, no studies are available in which comparison of the robustness of these two ELISA formats for food allergen detection is evaluated.

Chromatographic Techniques. Chromatographic techniques, with which direct detection of one or several allergenic proteins is possible, are also frequently used. For example, high-performance liquid chromatography (HPLC) methods with fluorometric detectors were developed for the detection of lysozyme, a known food allergen from hen’s egg, in dairy products.^{33,34} More complex protein mixtures, such as soybean

proteins in adulterated meat and bakery and dairy products could also be analyzed by HPLC.^{35–38} A more extensive review of liquid chromatographic (LC) methods developed for especially major food allergen detection is available elsewhere.³⁹ Nevertheless, the use of chromatographic methods coupled solely to UV or fluorescent detection can be troublesome in foods because the identification of the allergenic proteins can be hampered by the presence of other matrix proteins that can coelute and might lead to false-positive results. Therefore, coupling liquid chromatographic equipment to mass spectrometric analyzers enables the unambiguous confirmation of the presence of the allergens.

Mass Spectrometry (MS). MS techniques are extensively used to study proteins and peptides. Because food allergens are mostly proteins, many MS methods for their identification and detection are reported as well. Most of the reported methods involve separation via SDS-PAGE, 2D-PAGE, or LC prior analysis by MS of the either intact protein or peptides obtained after proteolytic digestion. Detection of intact allergenic proteins, especially derived from milk, was previously reported.^{40–42} Techniques in which detection of food allergens is based on the finding of specific markers resulting from the proteolytic digestion of native and/or modified proteins are also often employed. When using such approaches, it is important to identify stable peptide markers, especially in the development of quantitative methods. Targeted detection of such selected molecules (selected reaction monitoring) or multireaction monitoring enables high selectivity and thus accurate quantification. So far, peptide markers for peanuts were identified and used for the development of quantitative MS techniques.^{43–46} A more extensive review of mass spectrometric techniques used for the identification and detection of food allergen detection is available elsewhere.^{5,18,39}

The mass spectrometric methods represent a valuable tool for the confirmation of the presence of food allergens in different food matrices if contradictory results are obtained using other methods. Unfortunately, due to the fact that the equipment used is rather expensive and highly qualified personnel are needed to operate such systems, the MS techniques are not appropriate for use in routine analysis in a food company based setting, contrasting typically with immunological-based methods.

DNA-Based Methods. The principle of the DNA-based methods involves targeting a segment of the gene coding for the allergenic or other proteins of interest and amplifying only this DNA fragment to make it detectable.¹⁸ Unfortunately, DNA-based methods do not detect the allergen itself, and the results are difficult to correlate to actual allergen quantities and thus cannot be used in a proper risk assessment and management. Despite these disadvantages, DNA methods are extensively used for allergen detection because of their ease in application.^{47–52}

Challenges Related to Allergen Detection in Food.

The main disadvantage of the receptor-based method is the fact that small variations in the analytical target lead to great variability of the analytical outcome. De Meulenaer et al.⁵³ showed that antibodies developed toward roasted Virginia peanuts had a cross-reactivity of below 50% with other roasted peanuts varieties. This indicates that underestimations of the actual contamination levels might occur. A certain level of cross-reactivity, especially when different varieties of allergenic commodities are analyzed, is advisable. On the other hand, cross-reactivity with other food components (especially bulk

proteins) might lead to false-positive results or overestimation of the contamination levels. For example, if antibodies raised against hazelnut proteins have 1% cross-reactivity with milk proteins, this would lead to an analytical result of 100 ppm hazelnut in a cookie containing 1% milk proteins when extraction is performed on 1 g cookie/10 mL extraction buffer. Additionally, major issues related to extraction, matrix effects, and impact of processing can be expected during food allergen analysis. Therefore, they are further discussed in more detail.

Incomparable Results with Commercial Kits. Unfortunately, there is no general agreement on the expression of reporting units, which makes it difficult to compare the results of allergen detection kits. Some kits use standards reported as “amount of allergenic commodity”, whereas others report “amount allergenic protein”. Because the protein contents in different commodities (soybean, tree nuts, lupine seeds, etc.) might depend on the variety, this hampers comparison of the results between kits. A similar issue is valid in the interpretation of results of LC-MS-based methods. Indeed, typically one or several particular indicator peptides are used to quantify the content of a particular allergenic commodity in the analyzed food, based on the concentration of the indicator peptide in a kind of reference allergenic commodity. Again, the concentration of such an indicator peptide may depend upon several factors, however, and cannot be regarded as absolutely constant.⁵⁴ Coming back to immune assays, we have shown in a previous study that detection of native hazelnut and soybean proteins depends highly on the type of commercial kit used^{55,56} (Tables 2 and 3). When four commercial kits for

Table 2. Ratio (Percent) of Measured over Actual Hazelnut Protein Concentration after Duplicate Analyses of Hazelnut Proteins (40 ng/mL) either in Native Form or after 48 h of Heat Treatment in the Absence or Presence of Glucose⁵⁵

	Veratox	Ridascreen	HN residue	Biokits
native hazelnut	5.32	85.69	73.08	116.54
heat-treated hazelnut	4.33	79.76	61.13	85.23
heat-treated hazelnut with glucose	1.80	112.15	39.99	59.26

Table 3. Ratio (Percent) of Measured over Actual Soybean Protein Concentration after Duplicate Analyses of Soybean Proteins (50 ng/mL) either in Native Form or after 48 h of Heat Treatment in the Absence or Presence of Glucose⁵⁶

	Veratox	soy residue	Biokits
native soybean	374.67	84.81	<i>a</i>
heat-treated soybean	nd ^b	nd	364.72
heat-treated soybean with glucose	nd	nd	176.67

^aNot calculated; absorbance value obtained for the present concentration was above the calibration range. ^bnd, not detected.

hazelnut were tested, only one kit proved to give accurate results with untreated hazelnut proteins, two kits underestimated the actual protein content by on average 20%, and another kit underestimated the actual content by over 90%. On the contrary, one of the commercial kits for soybean highly overestimated the actual protein content and another underestimated by about 20%. When heat-treated samples were analyzed in either the presence or absence of glucose, the obtained results were also kit-dependent, although mostly an underestimation was observed. It should be noted that these

results were obtained using relatively simple protein solutions, not real food matrices. Furthermore, the performance of commercial kits depends highly on the type of food matrix analyzed as well. Whitaker et al.,⁵⁷ for instance, showed that the performance of four commercial peanut kits was especially poor in cookies.

The accuracy of commercial kits depends not only upon the chemical modifications induced in the untreated food but also upon the actual protein concentration present.^{56–59} As a consequence, care should be taken when using such kits for quantitative analysis because erroneous decisions related to risk assessment can be made.

Matrix Effects. Another important issue related to the detection of food allergens, by any of the above-mentioned methods, is that they are present in trace amounts and their presence might be masked by the matrix. Especially the receptor-based methods tend to perform differently depending on the type of matrix analyzed. This is mostly caused by (i) interaction of the analyte with the matrix, which hinders its extraction, or (ii) coextraction of matrix proteins, which can nonspecifically bind with antibodies, therefore giving false-positive results. However, hindered extractability due to interaction of analyte with the matrix can affect the detectability of food allergens by MS methods as well.^{60,61} Interaction with matrix components, such as polyphenols and tannins from chocolate, might impair the extractability of the analyte.⁶²

Husain et al.⁶³ showed that extracts from foods can influence the IC_{50} value of the calibration curves at different degrees. To take into account such matrix effects, it is possible to work with calibration curves prepared in extracts of particular allergen-free foods.^{63–65} Unfortunately, this means that the developed methods should be validated for each particular food product separately.

Processing. Food allergens (proteins) have a very complex structure, and upon processing they can be affected through numerous ways. They can be denatured with disruption of the tertiary and secondary structure, which might lead to modification of the conformational epitopes; they can be modified through Maillard reaction or partial hydrolysis, which might modify the linear epitopes; and they can aggregate and lose solubility.

As previously mentioned, ELISA methods are based on the molecular recognition between the receptor (antibody) and the analytical target (the proteins). However, due to processing the interaction between the antibodies and the modified allergens can be affected, which can lead to erroneous results. Unfortunately, a decreased detectability due to allergen modifications does not necessarily mean a decrease in allergenicity, with, in some cases, even an increase in allergenicity being observed.^{55,66–70} It is therefore of outmost importance to have reliable methods able to detect not only the native allergens but also the allergens modified through reactions typically occurring during processing and storage.

Previously, it was shown that detection of proteins modified through the Maillard reaction, protein oxidation, and partial hydrolysis is severely affected if commercial kits are used.^{58,59,71–77} Complete loss of detectability of an in-house-developed method using antibodies against Kunitz trypsin inhibitors was observed in cookies prepared with lactose,⁶⁴ whereas Ecker et al.⁷⁸ observed a decrease in detectability of lupine in cookies upon baking. Additionally, losses of detectability using LC-MS in strongly processed dairy products were also reported,⁴⁰ indicating that MS-based proteomic

approaches are prone to protein modifications as well. Such loss in detectability might be either due to the fact that the targeted proteins and/or peptides are modified during processing, resulting in mass shifts and poor ionization, or because of reduced proteolysis.⁷⁹ The integrity of DNA can also be modified during processing, which might lead to erroneous results as well.^{73,80,81}

Extractability. Extraction represents another important cause of erroneous results obtained by all of the analytical methods used. Any of the analytical methods mentioned above will only detect what is extracted. The yield of the extracted allergen depends on the type of allergen analyzed and the degree of modifications induced by processing. This means that the extraction methods should be also validated for specific products and processing conditions to help evaluate their applicability. Thermal processing impairs the solubility of the allergens,⁸² and this can directly affect the robustness of the developed methods, most often leading to false-negative results. Fu et al.⁷⁴ showed that dry or moist heating of whole egg powder decreased with over 75% of the yield of extractable protein content. Similarly, Monaci et al.⁷⁵ reported an over 80% decrease in the yield of the extractable proteins from cookies baked for 9 min at 180 °C. It is therefore important to make sure that the maximum amount of the targeted analyte is extracted.

Furthermore, when new analytical methods for food allergen detection are developed, the robustness is often evaluated on the basis of the determination of the recovery after spiking allergen-free products shortly before the extraction step.^{63,83–88} Even in such cases the time when the spiking is performed is critical. A comparison of the recovery assessed on samples spiked 24 h before extraction revealed that the recovery is significantly lowered, probably due to interactions with the matrix compared to spiking shortly before extraction even if no processing is applied.³³ Similarly, good recoveries were obtained when blank cookies were spiked with hazelnut shortly before extraction (73–107%).⁶⁵ On the contrary, spiking 2 h before extraction led to an approximately 60% decrease in recovery, again without the application of any processing step as such. However, contamination of food products with allergens is most likely caused by the use of contaminated primary materials or due to the use of shared equipment, and this before any food-processing conditions are applied. Therefore, determination of the recovery in samples incurred before the application of any processing is more accurate and should be applied in a proper validation process of any analytical method. Several studies in which evaluation of the robustness by incurring samples before processing was done are published.^{64,89–96} We have shown that when determining the robustness in cookies incurred before processing was applied, only about 10% recovery were obtained.⁶⁵ Similarly, the recovery of the DNA seems to also be affected by processing, mainly because of its degradation and/or hindered extractability.⁵¹

In conclusion, all of the analytical methods mentioned above are prone to erroneous results, especially if the extractability of the analyte is reduced. Obviously, the results obtained by all methods applied should always be considered with outmost care. False-negative results can present a potentially fatal risk for allergic consumers, whereas false-positive results may lead to unnecessary product withdrawal. It is therefore advisable to be very critical when using such methods because the results obtained are mostly semiquantitative or qualitative but, in

general, unreliable. Ideally, these methods should be validated via a case-to-case approach (individual allergens and as many matrices as possible), but due to the number of possible products produced within a single plant, this is not realistic.

■ NEW TRENDS IN THE QUANTIFICATION OF FOOD ALLERGENS

To overcome the above-mentioned issues related to the detection of food allergens in processed foods, new strategies for the development of more robust methods can be employed.

Use of Antibodies Raised against Modified Proteins.

Most of the ELISA methods developed for food allergen detection use antibodies raised against one or several nonmodified proteins (protein extracts).^{63,78,86,88,92,94,97–102} However, as above-mentioned, decreases in detectability of food allergens due to hampered recognition of the modified analytical target by the used antibodies can occur. Therefore, development of antibodies against proteins modified through reactions typically occurring during processing might help improve the detectability. We have previously shown that the detectability of soybean proteins could be improved by using antibodies against modified soybean protein extract compared to antibodies raised against Kunitz trypsin inhibitor in especially highly processed foods.⁶⁴ In general, antibodies can be raised either against thermally treated protein extracts^{64,65} or protein extracts from processed foods^{53,83,87,91,103–105} or against partially hydrolyzed proteins.¹⁰⁶ Gaskin et al.⁹¹ developed an ELISA method in which antibodies raised against protein extracts from raw and roasted cashews were used. Using antibodies with good affinity to the native as well as modified proteins can help to improve the detectability of allergens in unprocessed and processed foods.

Use of Antibodies Raised against Stable Proteins.

Using antibodies raised against one stable protein could also be a potential strategy to improve the robustness of analytical assays. In such cases it is crucial to investigate how stable the proteins are under different processing conditions. In a previous study we have shown that Cor a 9 from hazelnut is rather stable upon incubation with carbohydrates and lipids.⁵⁸ Moreover, several stable peptides derived from Cor a 9 could be detected in modified hazelnut protein extract as well. An additional advantage of choosing the Cor a 9 as analytical target is its abundance in hazelnut; thus, it can be more easily detected in foodstuffs contaminated by hazelnuts. Therefore, Cor a 9 was evaluated as a promising target for the detection of hazelnut traces in processed foods. When using anti-Cor a 9 antibodies in a sandwich ELISA, the robustness of the receptor-based analytical methodology was improved.²⁶ Similarly, antibodies against a single allergen such as the stable Gly m Bd 30K allergen were used for the detection of soybean^{93,95} and antibodies against the 2S albumin for the detection of walnut in processed foods.⁹⁶

Use of Stable Peptides as Analytical Targets. A relatively new strategy employed to improve the detectability of food allergens via receptor-based methods is by using antibodies raised against a single peptide. Earlier, Akkerdaas et al.¹⁰⁶ showed that using an ELISA with antibodies raised against pepsin-digested hazelnut proteins can improve detectability of hazelnut residues in chocolate. You et al.¹⁰⁷ and Liu et al.¹⁰⁸ developed antibodies against specific epitopes from β -conglycinin. When using such approaches, it is of utmost importance to select peptides that are stable during processing, ensuring that the affinity of the antibody toward the selected

peptide is not affected. On the other hand, stable peptides are also needed for the development of accurate quantitative MS techniques. As mentioned previously, marker peptides were identified only for a limited number of food allergens. In a previous study, we have identified stable peptides derived from whey, soybean, and hazelnut allergens.^{109–111}

Multiallergen Methods. Multiallergen detection methods are often used for diagnosis of allergy.^{112,113} However, interest in the development of receptor-based methods as well as mass spectrometric techniques for the detection of several allergens in foods is growing rapidly. Rejeb et al.¹⁰⁴ developed a multiallergen immunoassay for the detection of several nuts. Multiallergen mass spectrometric methods were developed as well. Heick et al.¹¹⁴ developed an LC-MS method that allowed the simultaneous detection of seven allergens (milk, egg, soybean, hazelnut, peanut, walnut, and almond). Bignardi et al.¹¹⁵ developed a multiallergen MS method for the detection of several tree nuts (cashew, hazelnut, almond, peanut, and walnut) with detection limits of as low as 10 mg/kg. Tortajada-Genaro et al.¹¹⁶ developed a DNA microarray method for the simultaneous detection of hazelnut, peanut, and soybean in foods, whereas Ehlert et al.¹¹⁷ developed a PCR method for the simultaneous detection of 10 allergens (peanuts, cashews, pecans, pistachios, hazelnuts, sesame seeds, macadamia nuts, almonds, walnuts, and Brazil nuts). Development of such multiallergen detection methods is interesting and can be especially useful for the screening of incoming ingredients from different suppliers and also for the fast screening of the final products.

Disregarding these new strategies that can be used to improve the detectability of allergens, the number of variables that can affect their quantitative detection is enormous. Therefore, it is difficult to say with certitude whether a certain food product is free of allergens or not. In the absence of a reliable and validated extraction and/or detection method, food producers should take the responsibility for producing allergen-free products through a proper allergen risk management.

■ ALLERGEN RISK MANAGEMENT

The three important quality assurance systems for a food company to ensure food safety and hygiene are¹¹⁸ GMP (Good Manufacturing Practice), PRP (Prerequisite Programs), and HACCP (Hazard Analysis Critical Control Points). Within this approach, a PRP is foreseen for allergen risk management on a company level. Allergen risk management does not have the intention to make a whole new food safety management system, but it should be included as a basic condition in a food company to control and manage direct allergens and hidden allergens due to cross-contaminations.¹¹⁹ The main objective of allergen risk management must be to prevent adverse reactions in food-allergic consumers without unnecessarily limiting their food choices. Extraction and analytical methods are necessary to back up the allergen risk management in validation and verification activities, for example, screening of raw materials and ingredients and validation of the impact of preventive measures such as cleaning or separate storage activities.

Food companies are responsible for the safety of food products brought onto the market and must implement a food safety management system based on good practices and HACCP principles.¹⁶ Allergen risk management means that the chance for the inadvertent presence of allergens in the end product through, for example, cross-contamination, is thoroughly evaluated. Several guidelines are available for the food

industry to set up an allergen management (e.g., on international level VITAL via <http://www.allergenbureau.net/> or for Europe via <http://www.eu-vital.org/en/home.html>). Many of them are obviously very generic, and a further tailoring to the company-specific setting and validation is therefore necessary. In general, the food industry should evaluate particular available methods (either commercial or in-house developed) for their reliability within the company, and when these prove to be so, then the allergen management program can be validated. This especially because some tests can prove to be extremely accurate with particular food matrix and processing conditions, but not with others, as earlier also discussed.

Several key points of the production process need to be considered and managed (i.e., product design and formulation, raw material purchase, cross-contamination during production process, evaluation of the impact of rework, labeling, and finally validation and verification). Figure 1 illustrates the concept of

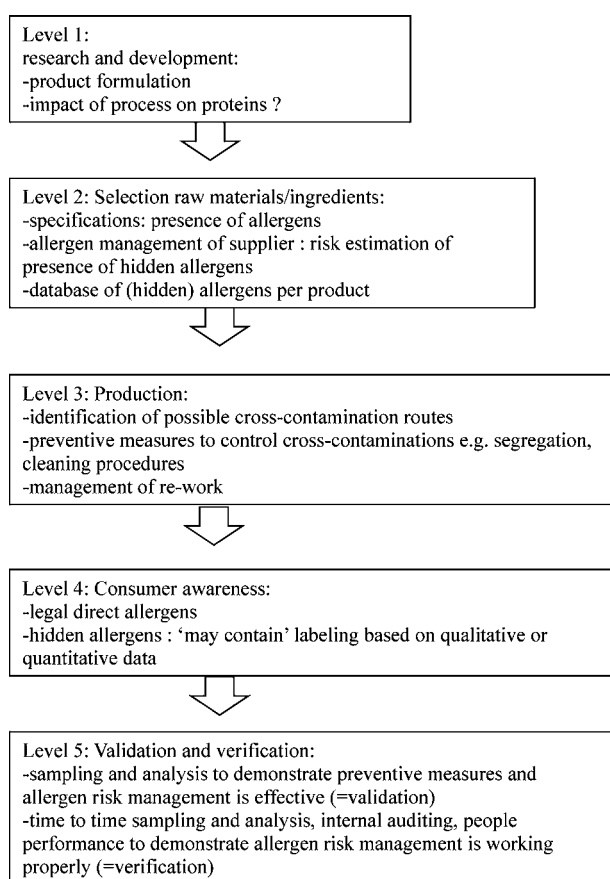


Figure 1. Overview of the concept of allergen risk management.

allergen risk management on the company level. Ideally, the complete physical separation of allergen-free and allergen-containing production lines or areas should solve the cross-contamination issues. However, this approach is mostly not economically feasible for the producer, and despite this, allergens could still be introduced in the plants when contaminated raw materials or ingredients are purchased. The supplier must provide accurate information to the producer concerning the presence of allergens in the raw materials or ingredients. Also, information about possible cross-contamination should be transferred to the producer. Forms or

checklists can be used to give the correct information.¹²⁰ When a new food product is developed, it is important to determine whether the allergens used in the product can be replaced by a nonallergic component.¹²⁰ When using new ingredients, food business operators should always determine whether they contain allergens or not. A change of the production process could also affect the final presence of allergens in a food product.¹²¹

Unfortunately, because “zero tolerance” level is difficult to achieve, preventive measures should be taken to avoid cross-contamination. Little is known about the frequency or amount of cross-contamination of allergens due to shared materials or infrastructures. Cross-contamination can occur through several ways: air, contact materials, via personnel, carry-over from batches, and through media such as oil and water during reuse.¹²² An important preventive measure is cleaning. Validation of the cleaning is necessary to estimate the risk of cross-contamination correctly and to adjust the cleaning methods.¹²³ Roder et al.¹³ investigated the cleaning efficiency for the reduction of cross-contamination of hazelnuts in the industrial production of cookies. Kerkaert et al.¹²² used lysozyme as an indicator for protein carry-over in fresh-cut vegetables through washing water. They reported that allergenic proteins can be transferred via wash water to the fresh-cut vegetables in the next batch to quantities which are able to pose a risk for the allergic consumers. This innovative approach could potentially be extended to validate, for example, cleaning or other typical processes. By using one or several indicator proteins of which the analytics are properly validated with respect to the actual process in which they are applied, their carry-over can be monitored in a quantitative way, thus enabling extrapolation of their behavior to that of a particular allergen present in the product as well. The rework of products represents another significant potential risk. Using this knowledge in combination with the accurate control of the critical points in production lines, prevention of contamination of allergen-free foods might be facilitated. Furthermore, it is important that products containing allergens are clearly labeled and kept separate if they are later reused in other products.¹²⁰

Unfortunately, because of the complexity of the food allergen issue, a single solution will probably not be realistic. Multiple strategies to support a validated allergen management on a company or even production line level seem to be the only valid approach to enable the food industry to produce safe foods for allergic patients.

Also at governmental level and the competent authority, inspecting and monitoring the safety of the food chain could progress toward other strategies to manage the problem of allergens. During inspections and audits performed in food companies, attention could be given toward the implementation of risk management at the company level, as previously described. Also during the monitoring of products available on the market, the reliability of used extraction/detection methods is questionable considering the issues raised earlier. A possible approach in monitoring and selecting the best extraction and detection method could be the evaluation of the impact of the process and matrix on the extraction and detection of the allergen from the food matrix. If a low impact is seen, then the methods can be applied for the monitoring purposes. However, from the discussion above it is clear that this is not always the case. Therefore, it would be recommended to know the recovery of the extraction and to know the best performing detection method for a certain food matrix. The obtained result

of the quantity of specific allergen could be recalculated for the loss of recovery toward the final food product. Instead of elaborating such an approach for each food product, a classification of food products could be made. This approach is in line with current European legislation for the migration of components from plastic food contact materials, in which reduction and correction factors are introduced to recalculate the actual migration from the obtained laboratory results for specific food products.¹²⁴

AUTHOR INFORMATION

Corresponding Author

*Phone: +32 92 64 61 66; Fax: + 32 92 64 62 15. E-mail: Bruno.DeMeulenaer@UGent.be.

Funding

We thank the Belgian Science Policy (SD/AF/03A) for financial support.

Notes

The authors declare no competing financial interest.

REFERENCES

- (1) Perry, T. T.; Scurlock, A. M.; Jones, S. M. Clinical manifestations of food allergic disease. In *Food Allergy*; Maleki, S., Burks, A. W., Helm, R. M., Eds.; ASM Press: Washington, DC, 2006; pp 3–17.
- (2) Sicherer, S. H.; Sampson, H. A. Food allergy. *J. Allergy Clin. Immunol.* **2006**, *117* (2), S470–S475.
- (3) Poms, R. E.; Klein, C. L.; Anklam, E. Methods for allergen analysis in food: a review. *Food Addit. Contam.* **2004**, *21* (1), 1–31.
- (4) Mills, E. N. C.; Mackie, A. R.; Burney, P.; Beyers, K.; Frewer, L.; Madsen, C.; Botjes, E.; Crevel, R. W. R.; van Ree, R. The prevalence, cost and basis of food allergy across Europe. *Allergy* **2007**, *62* (7), 717–722.
- (5) Monaci, L.; Visconti, A. Mass spectrometry-based proteomics methods for analysis of food allergens. *TrAC–Trends Anal. Chem.* **2009**, *28* (5), 581–591.
- (6) Cochrane, S.; Salt, L. J.; Wantling, E.; Rogers, A.; Coutts, J.; Ballmer-Weber, B. K.; Fritsche, P.; Fernandez, M. I.; Reig, I.; Knulst, A.; Le, T. M.; Asero, R.; Beyers, K.; Golding, M.; Crevel, R.; Mills, E. N. C.; Mackie, A. R. Development of a standardized low-dose double-blind placebo-controlled challenge vehicle for the EuroPrevall project. *Allergy* **2012**, *67* (1), 107–113.
- (7) Bindslev-Jensen, C.; Briggs, D.; Osterballe, M. Can we determine a threshold level for allergenic foods by statistical analysis of published data in the literature? *Allergy* **2002**, *57* (8), 741–746.
- (8) Van Putten, M. C.; Frewer, L. J.; Gilissen, L. J. W. J.; Gremmen, B.; Peijnenburg, A. A. C. M.; Wichers, H. J. Novel foods and food allergies: a review of the issues. *Trends Food Sci. Technol.* **2006**, *17* (6), 289–299.
- (9) European Parliament and Council Directive 2007/68/EC of 27 November 2007 amending Directive 2000/13/EC as regards certain food ingredients. *Off. J. Eur. Communities* **2007**, L310/11.
- (10) *Australia and New Zealand Food Standards Code Std 1.2.3; Mandatory warning and advisory statements and declarations*, 2011; <http://www.comlaw.gov.au/Series/F2008B00603>.
- (11) *Health Canada, Food Allergen Labelling*, 2012; <http://www.hc-sc.gc.ca/fn-an/label-etiquet/allergen/index-eng.php>.
- (12) The Food Allergen Labeling and Consumer Protection Act of 2004 (FALCPA); Washington, DC.
- (13) Roder, M.; Ibach, A.; Baltruweit, I.; Gruyters, H.; Janise, A.; Suwelack, C.; Matissek, R.; Vieths, S.; Holzhauser, T. Pilot plant investigations on cleaning efficiencies to reduce hazelnut cross-contamination in industrial manufacture of cookies. *J. Food Prot.* **2008**, *71* (11), 2263–2271.
- (14) *RASFF Rapid Alert System for Food and Feed*; European Commission: Brussels, Belgium, 2011.
- (15) Vierk, K.; Falci, K.; Wolyniak, C.; Klontz, K. C. Recalls of foods containing undeclared allergens reported to the US Food and Drug Administration, fiscal year 1999. *J. Allergy Clin. Immunol.* **2002**, *109* (6), 1022–1026.
- (16) EU Regulation 178/2002 Regulation (EC) No 178/2002 of the European Parliament and of the Council of 28 January 2002 laying down the general principles and requirements of food law, establishing the European Food Safety Authority and laying down procedures in matters of food safety. *Off. J. Eur. Communities* **2002**, 31.
- (17) Pele, M.; Brohée, M.; Anklam, E.; Van Hengel, A. J. Peanut and hazelnut traces in cookies and chocolates: relationship between analytical results and declaration of food allergens on product labels. *Food Addit. Contam.* **2007**, *24* (12), 1334–1344.
- (18) Kirsch, S.; Fourdrilis, S.; Dobson, R.; Scippo, M. L.; Maghain-Rogister, G.; De Pauw, E. Quantitative methods for food allergens: a review. *Anal. Bioanal. Chem.* **2009**, *395* (1), 57–67.
- (19) Schubert-Ullrich, P.; Rudolf, J.; Ansari, P.; Galler, B.; Fuhrer, M.; Molinelli, A.; Baumgartner, S. Commercialized rapid immunoanalytical tests for determination of allergenic food proteins: an overview. *Anal. Bioanal. Chem.* **2009**, *395* (1), 69–81.
- (20) Van Hengel, A. J. Food allergen detection methods and the challenge to protect food-allergic consumers. *Anal. Bioanal. Chem.* **2007**, *389* (1), 111–118.
- (21) Bremer, M. G. E. G.; Smits, N. G. E.; Haasnoot, W. Biosensor immunoassay for traces of hazelnut protein in olive oil. *Anal. Bioanal. Chem.* **2009**, *395* (1), 119–126.
- (22) Maier, I.; Morgan, M. R. A.; Lindner, W.; Pittner, F. Optical resonance-enhanced absorption-based near-field immunochip biosensor for allergen detection. *Anal. Chem.* **2008**, *80* (8), 2694–2703.
- (23) Mohammed, I.; Mullett, W. M.; Lai, E. P. C.; Yeung, J. M. Is biosensor a viable method for food allergen detection? *Anal. Chim. Acta* **2001**, *444* (1), 97–102.
- (24) Pollet, J.; Delpont, F.; Janssen, K. P. F.; Tran, D. T.; Wouters, J.; Verbiest, T.; Lammertyn, J. Fast and accurate peanut allergen detection with nanobead enhanced optical fiber SPR biosensor. *Talanta* **2011**, *83* (5), 1436–1441.
- (25) Raz, S. R.; Liu, H.; Norde, W.; Bremer, M. G. E. G. Food allergens profiling with an imaging surface plasmon resonance-based biosensor. *Anal. Chem.* **2010**, *82* (20), 8485–8491.
- (26) Trashin, S.; Cucu, T.; Devreese, B.; Adriaens, A.; De Meulenaer, B. Development of a highly sensitive and robust Cor a 9 specific enzyme-linked immunosorbent assay for the detection of hazelnut traces. *Anal. Chim. Acta* **2011**, *708*, 116–122.
- (27) Yman, I. M.; Eriksson, A.; Johansson, M. A.; Hellenas, K. E. Food allergen detection with biosensor immunoassays. *J. AOAC Int.* **2006**, *89* (3), 856–861.
- (28) Tran, D. T.; Janssen, K. P. F.; Pollet, J.; Lammertyn, E.; Anne, J.; Van Schepdael, A.; Lammertyn, J. Selection and characterization of DNA aptamers for egg white lysozyme. *Molecules* **2010**, *15* (3), 1127–1140.
- (29) Nadal, P.; Pinto, A.; Svobodova, M.; Canela, N.; O’Sullivan, C. K. DNA aptamers against the Lup a 1 food allergen. *PLoS One* **2012**, *7* (4).
- (30) *NEN-EN 15633 Foodstuffs – Detection of food allergens by immunological methods – Part 1: General considerations*; 2009.
- (31) *NEN-EN 15634 Foodstuffs – Detection of food allergens by molecular biological methods – Part 1: General considerations*; 2009.
- (32) *NEN EN 15842 Foodstuffs – Detection of food allergens – General consideration and validation of methods*; 2010.
- (33) Kerkaert, B.; Mestdagh, F.; De Meulenaer, B. Detection of hen’s egg white lysozyme in food: comparison between a sensitive HPLC and a commercial ELISA method. *Food Chem.* **2010**, *120* (2), 580–584.
- (34) Pellegrino, L.; Tirelli, A. A sensitive HPLC method to detect hen’s egg white lysozyme in milk and dairy products. *Int. Dairy J.* **2000**, *10* (7), 435–442.
- (35) Castro-Rubio, F.; Garcia, M. C.; Rodriguez, R.; Marina, M. L. Simple and inexpensive method for the reliable determination of additions of soybean proteins in heat-processed meat products: An

alternative to the AOAC official method. *J. Agric. Food Chem.* **2005**, *53* (2), 220–226.

(36) Garcia, M. C.; Dominguez, M.; Garcia-Ruiz, C.; Marina, M. L. Reversed-phase high-performance liquid chromatography applied to the determination of soybean proteins in commercial heat-processed meat products. *Anal. Chim. Acta* **2006**, *559* (2), 215–220.

(37) Garcia, M. C.; Marina, M. L. Rapid detection of the addition of soybean proteins to cheese and other dairy products by reversed-phase perfusion chromatography. *Food Addit. Contam.* **2006**, *23* (4), 339–347.

(38) Saz, J. M.; Marina, M. L. High performance liquid chromatography and capillary electrophoresis in the analysis of soybean proteins and peptides in foodstuffs. *J. Sep. Sci.* **2007**, *30* (4), 431–451.

(39) Faeste, C. K.; Ronning, H. T.; Christians, U.; Granum, P. E. Liquid chromatography and mass spectrometry in food allergen detection. *J. Food Prot.* **2011**, *74* (2), 316–345.

(40) Czerwenka, C.; Maier, I.; Potocnik, N.; Pittner, F.; Lindner, W. Absolute quantitation of beta-lactoglobulin by protein liquid chromatography - Mass spectrometry and its application to different milk products. *Anal. Chem.* **2007**, *79* (14), 5165–5172.

(41) Monaci, L.; van Hengel, A. J. Development of a method for the quantification of whey allergen traces in mixed-fruit juices based on liquid chromatography with mass spectrometric detection. *J. Chromatogr., A* **2008**, *1192* (1), 113–120.

(42) Weber, D.; Raymond, P.; Ben Rejeb, S.; Lau, B. Development of a liquid chromatography-tandem mass spectrometry method using capillary liquid chromatography and nano-electrospray ionization-quadrupole time-of-flight hybrid mass spectrometer for the detection of milk allergens. *J. Agric. Food Chem.* **2006**, *54* (5), 1604–1610.

(43) Careri, M.; Costa, A.; Elviri, L.; Lagos, J. B.; Mangia, A.; Terenghi, M.; Cereti, A.; Garoffo, L. P. Use of specific peptide biomarkers for quantitative confirmation of hidden allergenic peanut proteins Ara h 2 and Ara III 3/4 for food control by liquid chromatography-tandem mass spectrometry. *Anal. Bioanal. Chem.* **2007**, *389* (6), 1901–1907.

(44) Chassaing, H.; Norgaard, J. V.; van Hengel, A. J. Proteomics-based approach to detect and identify major allergens in processed peanuts by capillary LC-Q-TOF (MS/MS). *J. Agric. Food Chem.* **2007**, *55* (11), 4461–4473.

(45) Shefcheck, K. J.; Musser, S. M. Confirmation of the allergenic peanut protein, Ara h 1, in a model food matrix using liquid chromatography/tandem mass spectrometry (LC/MS/MS). *J. Agric. Food Chem.* **2004**, *52* (10), 2785–2790.

(46) Shefcheck, K. J.; Callahan, J. H.; Musser, S. M. Confirmation of peanut protein using peptide markers in dark chocolate using liquid chromatography-tandem mass spectrometry (LC-MS/MS). *J. Agric. Food Chem.* **2006**, *54* (21), 7953–7959.

(47) Costa, J.; Mafra, I.; Kuchta, T.; Oliveira, M. B. P. P. Single-tube nested real-time PCR as a new highly sensitive approach to trace hazelnut. *J. Agric. Food Chem.* **2012**, *60* (33), 8103–8110.

(48) Demmel, A.; Hupfer, C.; Busch, U.; Engel, K. H. Quantification of lupine (*Lupinus angustifolius*) in wheat flour using real-time PCR and an internal standard material. *Eur. Food Res. Technol.* **2012**, *235* (1), 61–66.

(49) Fuchs, M.; Cichna-Markl, M.; Hohegger, R. Development and validation of a novel real-time PCR method for the detection of celery (*Apium graveolens*) in food. *Food Chem.* **2012**, *130* (1), 189–195.

(50) Herrero, B.; Vieites, J. M.; Espineira, M. Fast real-time PCR for the detection of crustacean allergen in foods. *J. Agric. Food Chem.* **2012**, *60* (8), 1893–1897.

(51) Platteau, C.; De Loose, M.; De Meulenaer, B.; Taverniers, I. Detection of allergenic ingredients using real-time PCR: a case study on hazelnut (*Corylus avellana*) and soy (*Glycine max*). *J. Agric. Food Chem.* **2011**, *59* (20), 10803–10814.

(52) Lee, J. Y.; Kim, C. J. Determination of allergenic egg proteins in food by protein-, mass spectrometry-, and DNA-based methods. *J. AOAC Int.* **2010**, *93* (2), 462–477.

(53) De Meulenaer, B.; De la Court, M.; Acke, D.; De Meyere, T.; Van de Keere, A. Development of an enzyme-linked immunosorbent assay for peanut proteins using chicken immunoglobulins. *Food Agric. Immunol.* **2005**, *16* (2), 129–148.

(54) Weber, D.; Xie, R.; Cleroux, C.; Poirier, C.; Koerner, T. B. Hydrolyzed gluten and mass spectrometry: how much gluten is really present? *243rd ACS National Meeting & Exposition, AGDF Division*, San Diego, CA; American Chemical Society: Washington, DC, 2012; Abstr. 214.

(55) Cucu, T.; De Meulenaer, B.; Britts, C.; Devreese, B.; Ebo, D. Impact of thermal processing and the Maillard reaction on the basophil activation of hazelnut allergic patients. *Food Chem. Toxicol.* **2012**, *50* (5), 1722–1728.

(56) Platteau, C.; Cucu, T.; De Meulenaer, B.; Devreese, B.; De Loose, M.; Taverniers, I. Effect of protein glycation in the presence or absence of wheat proteins on detection of soybean proteins by commercial ELISA. *Food Addit. Contam.* **2011**, *28* (2), 127–135.

(57) Whitaker, T. B.; Williams, K. M.; Trucksess, M. W.; Slate, A. B. Immunochemical analytical methods for the determination of peanut proteins in foods. *J. AOAC Int.* **2005**, *88* (1), 161–174.

(58) Cucu, T.; Platteau, C.; Taverniers, I.; Devreese, B.; De Loose, M.; De Meulenaer, B. ELISA detection of hazelnut proteins: effect of protein glycation in the presence or absence of wheat proteins. *Food Addit. Contam.* **2011**, *28* (1), 1–10.

(59) Platteau, C. M. F.; Cucu, T.; Taverniers, I.; Devreese, B.; De Loose, M.; De Meulenaer, B. Effect of oxidation in the presence or absence of lipids on hazelnut and soybean protein detectability by commercial ELISA. *Food Agric. Immunol.* **2012**, DOI: doi.org/10.1080/09540105.2012.677009.

(60) Lutter, P.; Parisod, V.; Weymuth, H. Development and validation of a method for the quantification of milk proteins in food products based on liquid chromatography with mass spectrometric detection. *J. AOAC Int.* **2011**, *94* (4), 1043–1059.

(61) Monaci, L.; Losito, I.; Palmisano, F.; Visconti, A. Identification of allergenic milk proteins markers in fined white wines by capillary liquid chromatography-electrospray ionization-tandem mass spectrometry. *J. Chromatogr., A* **2010**, *1217* (26), 4300–4305.

(62) Vincent, D.; Wheatley, M. D.; Cramer, G. R. Optimization of protein extraction and solubilization for mature grape berry clusters. *Electrophoresis* **2006**, *27* (9), 1853–1865.

(63) Husain, F. T.; Bretbacher, I. E.; Nemes, A.; Cichna-Markl, M. Development and validation of an indirect competitive enzyme linked-immunosorbent assay for the determination of potentially allergenic sesame (*Sesamum indicum*) in food. *J. Agric. Food Chem.* **2010**, *58* (3), 1434–1441.

(64) Cucu, T.; Devreese, B.; Keshavarzian, A.; Rogge, M.; Vercruyse, L.; De Meulenaer, B. ELISA based detection of soybean proteins: a comparative study using antibodies against modified and native proteins. *Food Anal. Method.* **2012**, *5* (5), 1121–1130.

(65) Cucu, T.; Devreese, B.; Trashin, S.; Kerkaert, B.; Rogge, M.; De Meulenaer, B. Detection of hazelnut in foods using ELISA: challenges related to the detectability in processed foodstuffs. *J. AOAC Int.* **2012**, *95* (1), 149–156.

(66) Gruber, P.; Vieths, S.; Wangorsch, A.; Nerkamp, J.; Hofmann, T. Maillard reaction and enzymatic browning affect the allergenicity of Pru av 1, the major allergen from cherry (*Prunus avium*). *J. Agric. Food Chem.* **2004**, *52* (12), 4002–4007.

(67) Gruber, P.; Becker, W. M.; Hofmann, T. Influence of the Maillard reaction on the allergenicity of rAra h 2, a recombinant major allergen from peanut (*Arachis hypogaea*), its major epitopes, and peanut agglutinin. *J. Agric. Food Chem.* **2005**, *53* (6), 2289–2296.

(68) Maleki, S. J.; Chung, S. Y.; Champagne, E. T.; Raufman, J. P. The effects of roasting on the allergenic properties of peanut proteins. *J. Allergy Clin. Immunol.* **2000**, *106* (4), 763–768.

(69) Nakamura, A.; Watanabe, K.; Ojima, T.; Ahn, D. H.; Saeki, H. Effect of Maillard reaction on allergenicity of scallop tropomyosin. *J. Agric. Food Chem.* **2005**, *53* (19), 7559–7564.

(70) Nakamura, S.; Suzuki, Y.; Ishikawa, E.; Yakushi, T.; Jing, H.; Miyamoto, T. Reduction of in vitro allergenicity of buckwheat Fag e 1

through the Maillard-type glycosylation with polysaccharides. *Food Chem.* **2008**, *109* (3), 538–545.

(71) Downs, M. L.; Taylor, S. L. Effects of thermal processing on the enzyme-linked immunosorbent assay (ELISA) detection of milk residues in a model food matrix. *J. Agric. Food Chem.* **2010**, *58* (18), 10085–10091.

(72) Platteau, C.; Cucu, T.; De Meulenaer, B.; Devreese, B.; De Loose, M.; Taverniers, I. Effect of protein glycation in the presence or absence of wheat proteins on detection of soybean proteins by commercial ELISA. *Food Addit. Contam.* **2011**, *128* (2), 127–135.

(73) Scaravelli, E.; Brohee, M.; Marchelli, R.; van Hengel, A. J. The effect of heat treatment on the detection of peanut allergens as determined by ELISA and real-time PCR. *Anal. Bioanal. Chem.* **2009**, *395* (1), 127–137.

(74) Fu, T. J.; Maks, N.; Banaszewski, K. Effect of heat treatment on the quantitative detection of egg protein residues by commercial enzyme-linked immunosorbent assay test kits. *J. Agric. Food Chem.* **2010**, *58* (8), 4831–4838.

(75) Monaci, L.; Brohee, M.; Tregoeat, V.; van Hengel, A. J. Influence of baking time and matrix effects on the detection of milk allergens in cookie model food system by ELISA. *Food Chem.* **2011**, *127*, 669–675.

(76) Khuda, S.; Slate, A.; Pereira, M.; Al-Taher, F.; Jackson, L.; Diaz-Amigo, C.; Bigley, E. C.; Whitaker, T.; Williams, K. M. Effect of processing on recovery and variability associated with immunochemical analytical methods for multiple allergens in a single matrix: sugar Cookies. *J. Agric. Food Chem.* **2012**, *60* (17), 4195–4203.

(77) Khuda, S.; Slate, A.; Pereira, M.; Al-Taher, F.; Jackson, L.; Diaz-Amigo, C.; Bigley, E. C.; Whitaker, T.; Williams, K. M. Effect of processing on recovery and variability associated with immunochemical analytical methods for multiple allergens in a single matrix: dark chocolate. *J. Agric. Food Chem.* **2012**, *60* (17), 4204–4211.

(78) Ecker, C.; Cichna-Markl, M. Development and validation of a sandwich ELISA for the determination of potentially allergenic lupine in food. *Food Chem.* **2012**, *130* (3), 759–766.

(79) Johnson, P. E.; Baumgartner, S.; Aldick, T.; Bessant, C.; Giosafatto, V.; Heick, J.; Mamone, G.; O'Connor, G.; Poms, R. E.; Popping, B.; Reuter, A.; Ulberth, F.; Watson, A.; Monaci, L.; Mills, E. N. C. Current perspectives and recommendations for the development of mass spectrometry methods for the determination of allergens in foods. *J. AOAC Int.* **2011**, *94* (4), 1026–1033.

(80) Platteau, C.; De Loose, M.; De Meulenaer, B.; Taverniers, I. Quantitative detection of hazelnut (*Corylus avellana*) in cookies: ELISA versus real-time PCR. *J. Agric. Food Chem.* **2011**, *59* (21), 11395–11402.

(81) Kenk, M.; Panter, S.; Engler-Blum, G.; Bergemann, J. Sensitive DNA-based allergen detection depends on food matrix and DNA isolation method. *Eur. Food Res. Technol.* **2012**, *234* (2), 351–359.

(82) Schmitt, D. A.; Nesbit, J. B.; Hurlburt, B. K.; Cheng, H. P.; Maleki, S. J. Processing can alter the properties of peanut extract preparations. *J. Agric. Food Chem.* **2010**, *58* (2), 1138–1143.

(83) Drs, E.; Baumgartner, S.; Bremer, M.; Kemmers-Voncken, A.; Smits, N.; Haasnoot, W.; Banks, J.; Reece, P.; Danks, C.; Tomkies, V.; Immer, U.; Schmitt, K.; Krska, R. Detection of hidden hazelnut protein in food by IgY-based indirect competitive enzyme-immunoassay. *Anal. Chim. Acta* **2004**, *520* (1–2), 223–228.

(84) Faeste, C. K.; Plassen, C. Quantitative sandwich ELISA for the determination of fish in foods. *J. Immunol. Methods* **2008**, *329* (1–2), 45–55.

(85) Holzhauser, T.; Vieths, S. Quantitative sandwich ELISA for determination of traces of hazelnut (*Corylus avellana*) protein in complex food matrices. *J. Agric. Food Chem.* **1999**, *47* (10), 4209–4218.

(86) Holzhauser, T.; Vieths, S. Indirect competitive ELISA for determination of traces of peanut (*Arachis hypogaea* L.) protein in complex food matrices. *J. Agric. Food Chem.* **1999**, *47* (2), 603–611.

(87) Kiening, M.; Niessner, R.; Drs, E.; Baumgartner, S.; Krska, R.; Bremer, M.; Tomkies, V.; Reece, P.; Danks, C.; Immer, U.; Weller, M. G. Sandwich immunoassays for the determination of peanut and

hazelnut traces in foods. *J. Agric. Food Chem.* **2005**, *53* (9), 3321–3327.

(88) Ma, X.; Sun, P.; He, P. L.; Han, P. F.; Wang, J. J.; Qiao, S. Y.; Li, D. F. Development of monoclonal antibodies and a competitive ELISA detection method for glycinin, an allergen in soybean. *Food Chem.* **2010**, *121* (2), 546–551.

(89) de Luis, R.; Mata, L.; Estopanan, G.; Lavilla, M.; Sanchez, L.; Perez, M. D. Evaluation of indirect competitive and double antibody sandwich ELISA tests to determine β -lactoglobulin and ovomucoid in model processed foods. *Food Agric. Immunol.* **2008**, *19* (4), 339–350.

(90) de Luis, R.; Lavilla, M.; Sanchez, L.; Calvo, M.; Perez, M. D. Development and evaluation of two ELISA formats for the detection of β -lactoglobulin in model processed and commercial foods. *Food Control* **2009**, *20* (7), 643–647.

(91) Gaskin, F. E.; Taylor, S. L. Sandwich enzyme-linked immunosorbent assay (ELISA) for detection of cashew nut in foods. *J. Food Sci.* **2011**, *76* (9), T218–T226.

(92) Kaw, C. H.; Hefle, S. L.; Taylor, S. L. Sandwich enzyme-linked immunosorbent assay (ELISA) for the detection of lupine residues in foods. *J. Food Sci.* **2008**, *73* (8), T135–T140.

(93) Morishita, N.; Kamjya, K.; Matsumoto, T.; Sakai, S.; Teshima, R.; Urisu, A.; Moriyama, T.; Ogawa, T.; Akiyama, H.; Morimatsu, F. Reliable enzyme-linked immunosorbent assay for the determination of soybean proteins in processed foods. *J. Agric. Food Chem.* **2008**, *56* (16), 6818–6824.

(94) Reed, Z. H.; Park, J. W. Estimating the quantity of egg white and whey protein concentrate in prepared crabstick using ELISA. *Food Chem.* **2010**, *118* (3), 575–581.

(95) Sakai, S.; Adachi, R.; Akiyama, H.; Teshima, R.; Morishita, N.; Matsumoto, T.; Urisu, A. Enzyme-linked immunosorbent assay kit for the determination of soybean protein in processed foods: interlaboratory evaluation. *J. AOAC Int.* **2010**, *93* (1), 243–248.

(96) Sakai, S.; Adachi, R.; Akiyama, H.; Teshima, R.; Doi, H.; Shibata, H.; Urisu, A. Determination of walnut protein in processed foods by enzyme-linked immunosorbent assay: inter laboratory study. *J. AOAC Int.* **2010**, *93* (4), 1255–1261.

(97) Koppelman, S. J.; Knulst, A. C.; Koers, W. J.; Penninks, A. H.; Peppelman, H.; Vlooswijk, R.; Pigman, I.; van Duijn, G.; Hessing, M. Comparison of different immunochemical methods for the detection and quantification of hazelnut proteins in food products. *J. Immunol. Methods* **1999**, *229* (1–2), 107–120.

(98) Lee, P. W.; Hefle, S. L.; Taylor, S. L. Sandwich enzyme-linked immunosorbent assay (ELISA) for detection of mustard in foods. *J. Food Sci.* **2008**, *73* (4), T62–T68.

(99) Polenta, G.; Godefroy-Benrejeub, S.; Delahaut, P.; Weber, D.; Abbott, M. Development of a competitive ELISA for the detection of pecan (*Carya illinoensis*) traces in food. *Food Anal. Methods* **2010**, *3* (4), 375–381.

(100) Sletten, G. B.; Lovberg, K. E.; Moen, L. H.; Skarpeid, H. J.; Egaas, E. A comparison of time-resolved fluoroimmunoassay and ELISA in the detection of casein in foodstuffs. *Food Agric. Immunol.* **2005**, *16* (3–4), 235–243.

(101) Stumr, F.; Gabrovska, D.; Rvsova, J.; Hanak, P.; Plicka, J.; Tomkova, K.; Dvorska, P.; Cuhra, P.; Kubik, M.; Barsova, S.; Karsulinova, L.; Bulawova, H.; Brychta, J.; Yman, I. M. ELISA kit for casein determination: interlaboratory study. *J. AOAC Int.* **2010**, *93* (2), 676–682.

(102) Wang, H. Y.; Li, G.; Yuan, F.; Wu, Y. J.; Chen, Y. Detection of the allergenic celery protein component (Api g 1.01) in foods by immunoassay. *Eur. Food Res. Technol.* **2011**, *233* (6), 1023–1028.

(103) Holden, L.; Faeste, C. K.; Egaas, E. Quantitative sandwich ELISA for the determination of lupine (*Lupinus* spp.) in foods. *J. Agric. Food Chem.* **2005**, *53* (15), 5866–5871.

(104) Rejeb, S. B.; Abbott, M.; Davies, D.; Cleroux, C.; Delahaut, P. Multi-allergen screening immunoassay for the detection of protein markers of peanut and four tree nuts in chocolate. *Food Addit. Contam.* **2005**, *22* (8), 709–715.

(105) Scheibe, B.; Weiss, W.; Rueff, F.; Przybilla, B.; Gorg, A. Detection of trace amounts of hidden allergens: hazelnut and almond proteins in chocolate. *J. Chromatogr., B* **2001**, 756 (1–2), 229–237.

(106) Akkerdaas, J. H.; Wensing, M.; Knulst, A. C.; Stephan, O.; Hefle, S. L.; Aalberse, R. C.; van Ree, R. A novel approach for the detection of potentially hazardous pepsin stable hazelnut proteins as contaminants in chocolate-based food. *J. Agric. Food Chem.* **2004**, 52 (25), 7726–7731.

(107) You, J. M.; Li, D.; Qiao, S. Y.; Wang, Z. R.; He, P. L.; Ou, D. Y.; Dong, B. Development of a monoclonal antibody-based competitive ELISA for detection of β -conglycinin, an allergen from soybean. *Food Chem.* **2008**, 106 (1), 352–360.

(108) Liu, B.; Teng, D.; Yang, Y. L.; Wang, X. M.; Wang, J. H. Development of a competitive ELISA for the detection of soybean alpha subunit of β -conglycinin. *Process Biochem.* **2012**, 47 (2), 280–287.

(109) Cucu, T.; De Meulenaer, B.; Kerkaert, B.; Vandenberghe, I.; Devreese, B. MALDI based identification of whey protein derived tryptic marker peptides that resist protein glycation. *Food Res. Int.* **2012**, 47 (1), 23–30.

(110) Cucu, T.; De Meulenaer, B.; Devreese, B. MALDI based identification of soybean protein markers – possible analytical targets for allergen detection in processed foods. *Peptides* **2012**, 33 (2), 187–196.

(111) Cucu, T.; De Meulenaer, B.; Devreese, B. MALDI based identification of stable hazelnut protein derived tryptic marker peptides. *Food Addit. Contam.* **2012**, 29 (12), 1821–1831.

(112) Wuthrich, B.; Wydler, B. Allergy diagnosis: comparison of two multi-allergen dipstick screening tests (IgE quick and CMG Immunodot) between CAP FEIA and skin prick tests. *Allergologie* **1999**, 22 (4), 215–222.

(113) Deinhofer, K.; Sevcik, H.; Balic, N.; Harwanegg, C.; Hiller, R.; Rumpold, H.; Mueller, M. W.; Spitzauer, S. Microarrayed allergens for IgE profiling. *Methods* **2004**, 32 (3), 249–254.

(114) Heick, J.; Fischer, M.; Kerbach, S.; Tamm, U.; Popping, B. Application of a liquid chromatography tandem mass spectrometry method for the simultaneous detection of seven allergenic foods in flour and bread and comparison of the method with commercially available ELISA test kits. *J. AOAC Int.* **2011**, 94 (4), 1060–1068.

(115) Bignardi, C.; Elviri, L.; Penna, A.; Careri, M.; Mangia, A. Particle-packed column versus silica-based monolithic column for liquid chromatography-electrospray-linear ion trap-tandem mass spectrometry multiallergen trace analysis in foods. *J. Chromatogr., A* **2010**, 1217 (48), 7579–7585.

(116) Tortajada-Genaro, L. A.; Santiago-Felipe, S.; Morais, S.; Gabaldon, J. A.; Puchades, R.; Maquieira, A. Multiplex DNA detection of food allergens on a digital versatile disk. *J. Agric. Food Chem.* **2012**, 60 (1), 36–43.

(117) Ehlert, A.; Demmel, A.; Hupfer, C.; Busch, U.; Engel, K. H. Simultaneous detection of DNA from 10 food allergens by ligation-dependent probe amplification. *Food Addit. Contam.* **2009**, 26 (4), 409–418.

(118) Jacxsens, L.; Devlieghere, F.; Uyttendaele, M. *Quality Management Systems in the Food Industry*; Ghent University Press, 2009, 1–141.

(119) Crevel, R. Risk management – the principles. In *Management of Food Allergens*, 1st ed.; Coutts, J., Fielder, R., Eds.; Wiley-Blackwell: Oxford, UK, 2009; pp 83–101.

(120) Crevel, R. Industrial dimensions of food allergy. *Proc. Nutr. Soc.* **2005**, 64 (4), 470–474.

(121) Sathe, S. K.; Teuber, S. S.; Roux, K. H. Effects of food processing on the stability of food allergens. *Biotechnol. Adv.* **2005**, 23 (6), 423–429.

(122) Kerkaert, B.; Jacxsens, L.; Van De Perre, E.; De Meulenaer, B. Use of lysozyme as an indicator of protein cross-contact in fresh-cut vegetables via wash waters. *Food Res. Int.* **2012**, 45 (1), 39–44.

(123) Jackson, L. S.; Al Taher, F. M.; Moorman, M.; DeVries, J. W.; Tippett, R.; Swanson, K. M. J.; Fu, T. J.; Salter, R.; Dunaif, G.; Estes, S.; Albillos, S.; Gendel, S. M. Cleaning and other control and

validation strategies to prevent allergen cross-contact in food-processing operations. *J. Food Prot.* **2008**, 71 (2), 445–458.

(124) European Commission. . Commission Regulation (EU) No. 10/2011 on plastic materials and articles intended to come into contact with food. *Off. J. Eur. Union* **2011**, L12–1–L12/89.